

## Invivoscribe Products for SHM - RUO





## Gel & Capillary

IGH Somatic Hypermutation Assay v2.0
Gel Detection

IGH Somatic Hypermutation Assay v2.0
ABI Fluorescence Detection

#### NGS

LymphoTrack® IGHV Leader SHM Assay - MiSeq®

> LymphoTrack® IGH FR1 Assay – MiSeq®

> LymphoTrack® IGH FR1 Assay – S5/PGM™



# Invivoscribe Products for SHM - CE-IVD





### NGS

LymphoTrack® Dx IGHV Leader SHM Assay - MiSeq®

> LymphoTrack® Dx IGH FR1 Assay – MiSeq®

LymphoTrack® Dx IGH FR1 Assay – S5/PGM™



# **Invivoscribe Products for SHM**





	Catalog #								
RUO Products	Kit	MegaKit							
	(33 rxn)	(330 rxn)							
Gel	Gel Detection								
IGH Somatic Hypermutation Assay v2.0	5-101-0030	5-101-0040							
ABI Fluores	ABI Fluorescence Detection								
IGH Somatic Hypermutation Assay v2.0	5-101-0031	5-101-0041							

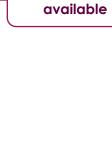


# **Invivoscribe RUO Products for SHM**





	Cata	log #		
RUO Products	Kit A 8 indices (40 rxn)	Panel 24 indices (120 rxn)		
	MiSeq <sup>®</sup>			
LymphoTrack <sup>®</sup> IGHV Leader Somatic Hypermutation Assay	7-121-0059	7-121-0069		
LymphoTrack <sup>®</sup> IGH FR1 Assay	7-121-0009	7-121-0039		
LymphoTrack <sup>®</sup> Software - MiSeq <sup>®</sup>	7-500-0009			
	12 indices (60 rxn)	Panel		
	S5/PGM <sup>TM</sup>			
LymphoTrack <sup>®</sup> IGHV Leader Somatic Hypermutation Assay	X	X		
LymphoTrack <sup>®</sup> IGH FR1 Assay	7-121-0007	X		
LymphoTrack® Software – S5/PGM <sup>TM</sup>	7-500-0007	X		



Up to 48 indices



# Invivoscribe CE-IVD Products for SHM





	Cata	log #			
CE-IVD Products	Kit A 8 indices (40 rxn)	Panel 24 indices (120 rxn)			
	MiSeq <sup>®</sup>				
LymphoTrack <sup>®</sup> Dx <i>IGHV</i> Leader Somatic Hypermutation Assay	9-121-0059	9-121-0069			
LymphoTrack <sup>®</sup> Dx <i>IGH</i> FR1 Assay	9-121-0009	9-121-0039			
LymphoTrack® Dx Software - MiSeq®	9-500-0009				
	12 indices (60 rxn)	Panel			
	S5/PGM <sup>TM</sup>				
LymphoTrack <sup>®</sup> Dx <i>IGHV</i> Leader Somatic Hypermutation Assay	X	Χ			
LymphoTrack® Dx IGH FR1 Assay	9-121-0007	X			
LymphoTrack® Dx Software – \$5/PGM <sup>TM</sup>	9-500-0007	X			



# LabPMM Clonality and SHM Testing





## **Services Catalog**

#### Available services include:

S CDx FLT3

NGS Gene Panels

Clonality testing (IGH, IGK, TRG & TRB)

MRD assays

Custom assays

Companion Diagnostic Tests	
Introduction	12
CDx - USA LeukoStrat® CDx <i>FLT3</i> Mutation Assay	14
CDx (CE-marked) LeukoStrat® CDx FLT3 Mutation Assay	16
CDx - Japan LeukoStrat® CDx FLT3 Mutation Assay	18
Molecular Diagnostic Tests	
Introduction NPM1 Mutation Analysis	20 22
Clonality NGS Tests	
Introduction  IGH Clonality Assays	24 26
IGH Somatic Hypermutation Assay	28
IGK Clonality Assay	30
TRB Clonality Assay TRG Clonality Assay	32 34
Minimal Residual Disease NGS Tests	
Introduction	36
AML <sup>1</sup>	
FLT3 ITD MRD Assay	38
NPM1 MRD Assays	40
Clonality <sup>2</sup>	***************************************
IGH MRD Clonality Assays	42
IGKMRD Clonality Assay	44
TRB MRD Clonality Assay	46
TRG MRD Clonality Assay	48
NGS Cancer Panels	
Introduction	50
MyAML®1	52
MyHEME®2	56
MyMRD®2	54







## Which options does Invivoscribe offer for SHM testing?

- Assays kits compatible with Gel, Capillary or NGS methods
- Controls
- Service
- All of the above



## Gel and Capillary assays for SHM analysis

IGH Somatic Hypermutation Assay v2.0 (RUO)





### **Kit Contents**

	Reagent Cor	mponents	Unit	Kit	MegaKit	
Reagent	Gel Detection	ABI Detection	Quantity	# of Units	# of Units	
	Hypermutation Mix 1 v2.0 - Unlabeled	Hypermutation Mix 1 v2.0 – 6-FAM	1500µL	1	10	
Master Mixes	Hypermutation Mix 2 v2.0 - Unlabeled	Hypermutation Mix 2 v2.0 – 6-FAM	1500µL	1	10	
Template Amplification Control Master Mix	Specimen Control Size Ladder - Unlabeled	Specimen Control Size Ladder – 6-FAM	1500µL	1	10	
Positive Control	IVS-0013 Clonal	Control DNA	100µL	1	5	
DNA and RNA	IVS-0013 Clonal	100µL	1	5		
Negative (Normal) Control DNA	IVS-0000 Polyclond	al Control DNA	100µL	1	5	
Soguencing Primer	IGH JH Primer -	Unlabeled	10µL	1	5	
Sequencing Primer	Primer – Hypermuta	tion - Unlabeled	10µL	1	5	





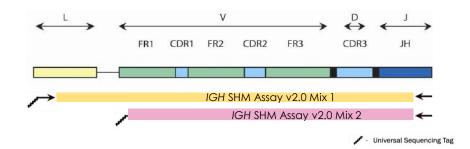
#### Gel and ABI Fluorescence Detection





- Contains forward primers that target the Leader (L) region
- Sequence upstream of the IGHV gene
- Allows a complete analysis of the IGHV gene

- Contains forward primers that target the Framework Region 1 (FR1)
- Allow an analysis of sequence between FR1 and the downstream joining (J) region of the IGHV gene

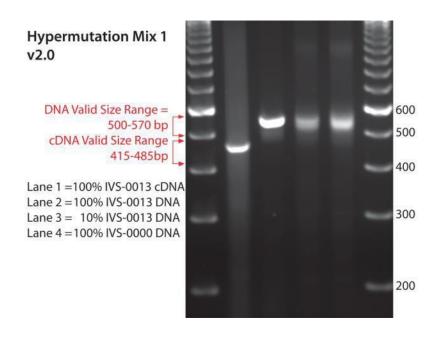






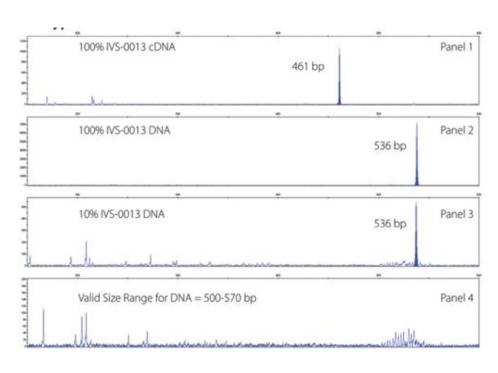
### Mix 1 Clonality Assessment

#### Gel Detection



- Mix 1 generates large product sizes which have low resolution when run on a gel
- For higher resolution, decrease voltage and run for a longer duration

#### **ABI Fluorescence Detection**



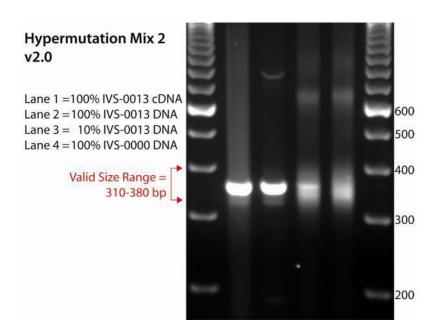
 The resolution is enhanced by the fluorescence label and peak detection software





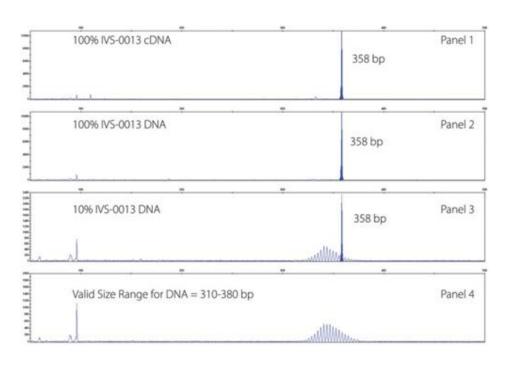
#### Mix 2 Clonality Assessment

#### Gel Detection



- Mix 2 generates smaller product sizes and has higher resolution, as compared to mix 1
- Mutation coverage by this primer set may be decreased relative to Mix 1 as products do not include the complete FR1 sequence

#### **ABI Fluorescence Detection**



 The resolution is enhanced by the fluorescence label and peak detection software

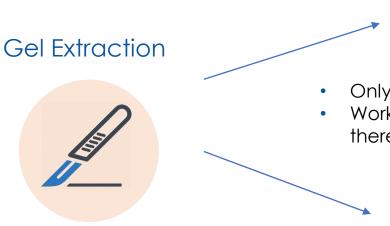




## Sequencing of PCR product

#### Cloning into a vector

#### Sanger Sequencing







- Only use with unlabeled amplicons
- Works best with weak clonal bands or if there is more than one clonal band



**Time Consuming and Labor Intensive** 

 Works best with weak clonal bands or if there is more than one clonal band

#### Direct Sanger Sequencing



- Works best with little to no background amplification and only one clonal product
- Risk of Sequencing Failures / Presence of multiple PCR products





### Data bank selection & Data analysis

#### Data Bank Selection

#### Sequence Data Analysis

#### Hypermutation Reporting







Find and align the germline V region sequence that best corresponds to the sample sequence.

Determine the number of mismatched bases and the total number of bases that are being compared.

% divergence = 
$$\frac{\text{N (mismatched bases)}}{\text{total N bases}}$$

% homology = 100% - % divergence

**IMGT** – The International ImMunoGeneTics information system

Analysis tools: IMGT/V-QUEST and IMGT/Junction Analysis

V BASE – The MRC Centre for Protein Engineering's Database of human antibody genes

Analysis tools: DNAPLOT

NCBI – National Center for Biotechnology Information

Analysis tool: IgBLAST (Basic Local Alignment Search Tool)

**Clonal Control**: Sequence data obtained from the positive IVS-0013 Clonal Control DNA or RNA should correspond to an unmutated, in-frame VH1-46 to JH4 rearrangement.

#### SHM status\*:

- % divergence ≥ 2%: Presence of IGH SHM
- % divergence <2%: Absence of IGH SHM</li>

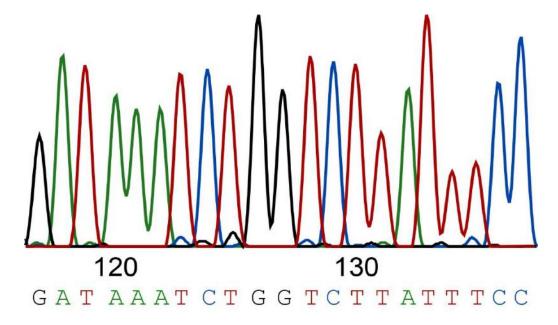


# Traditional Method for IGHV Analysis



## Sanger Sequencing is standard, **BUT** it has caveats

- May not differentiate different clonal populations in a sample
- Lower sensitivity
- Failure rate is 9-18%



Krober A, et al. Blood 2002; 100: 1410–1416. Austen B, et al. Blood 2005; 106: 3175–3182. Burger JA, et al. Lancet Oncol 2014; 15:1090–1099





# NGS Assays for SHM Analysis



# LymphoTrack® SHM Assays



## Widely Available Sequencing Platforms



### **NGS** Assays for SHM Analysis:

- LymphoTrack<sup>®</sup> IGHV Leader SHM Assay MiSeq<sup>®</sup>
- LymphoTrack<sup>®</sup> IGH FR1 Assay MiSeq<sup>®</sup>
- LymphoTrack<sup>®</sup> IGH FR1 Assay S5/PGM<sup>TM</sup>

Same kits as for Clonality Assessment!



# LymphoTrack® Dx SHM Assays



## Widely Available Sequencing Platforms



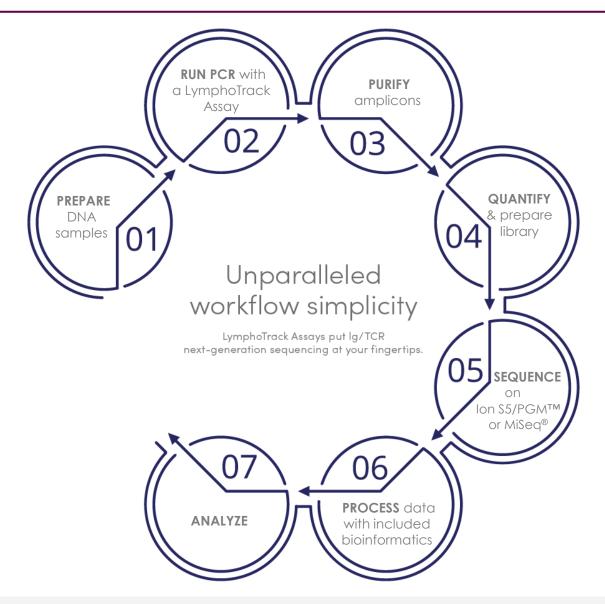
### **NGS** Assays for SHM Analysis:

- LymphoTrack® Dx IGHV Leader SHM Assay MiSeq®
- LymphoTrack® Dx IGH FR1 Assay MiSeq®
- LymphoTrack® Dx IGH FR1 Assay S5/PGM<sup>TM</sup>

Same kits as for Clonality Assessment!



## Same Assays and Workflow for Clonality & SHM





# Comprehensive Menu



Available Sequencing Platforms	MiSeq®	lon S5™	Ion PGM <sup>TM</sup>
Menu	B-Cell IGHV (Leader) IGH FR 1 IGH FR2 IGH FR3 IGK T-Cell TRG TRB	B-Cell  IGH FR1  IGH FR2  IGH FR3  IGK  T-Cell  TRG	B-Cell  IGH FR1  IGH FR2  IGH FR3  IGK  T-Cell  TRG
Kit size(s)	8-index or 24-index kits	12 barcodes	12 barcodes
Validated Sequencing Kits	V2 (2 x 150 bp) V2 (2 x 250 bp) V3 (2 x 300 bp)	Ion 520™ Ion 530™	lon 316™ v2 BC lon 318™ v2 BC

**SHM Analysis** 



# Comprehensive Menu

Available Sequencing Platforms	Miseq®	lon S5™	Ion PGM <sup>TM</sup>
Menu	B-Cell IGHV (Leader) IGH FR 1 IGH FR2 IGH FR3 IGK T-Cell TRG TRB	B-Cell  IGH FR 1  IGH FR 2  IGH FR 3  IGK  T-Cell  TRG	B-Cell  IGH FR1  IGH FR2  IGH FR3  IGK  T-Cell  TRG
Kit size(s)	8-index or 24-index kits	12 barcodes	12 barcodes
Validated Sequencing Kits	V2 (2 x 150 bp) V2 (2 x 250 bp) V3 (2 x 300 bp)	lon 520™ lon 530™	lon 316™ v2 BC lon 318™ v2 BC

**SHM Analysis** 



# LymphoTrack® SHM Assays



### **Kit Contents**

	Reagent Coi	mponents				
Reagent	IGHV Leader Hypermutation Somatic Assay - MiSeq®  IGH FR1 Assay - MiSeq®		Unit Quantity	Kit A # of Units	Panel # of Units	
		MiSeq <sup>®</sup>				
Master Mixes	IGH Leader MiSeq 08 indices	IGH FR1 MiSeq 08 indices	250	8	24	
Masiei Mixes	IGH Leader MiSeq 16 additional indices	<i>IGH</i> FR1 MiSeq 16 additional indices	- 250µl	8	24	
Positive	IGH SHM POS (+)	X				
Control DNA	IGH POS	S (+)	45µl	1	3	
Negative Control DNA	NGS NE	G (-)	- 40μι	'		
		S5/PGM <sup>TM</sup>				
Master Mixes	IGH FR1 S5 12 india		250µl	12	Х	
Positive Control DNA	IGH POS	S (+)	45l	2	V	
Negative Control DNA	NGS NE	G (-)	- 45µl	2	X	



# LymphoTrack® Dx SHM Assays

#### **Kit Contents**

	Reagent Cor	mponents				
Reagent	<i>IGHV</i> Leader Hypermutation Somatic Assay - MiSeq®	permutation Somatic IGH FRT Assay -		Kit A # of Units	Panel # of Units	
		MiSeq <sup>®</sup>				
Master Mixes	IGH Leader MiSeq 08 indices	IGH FR1 MiSeq 08 indices	250	8	24	
Masier Mixes	IGH Leader MiSeq 16 additional indices	<i>IGH</i> FR1 MiSeq 16 additional indices	- 250µl	8	24	
Positive	IGH SHM POS (+)	X				
Control DNA	IGH POS	S (+)	45µl	1	3	
Negative Control DNA	NGS NE	G (-)	τομι		· ·	
		S5/PGM <sup>TM</sup>				
Master Mixes	IGH FR1 S5 12 india		250µl	12	X	
Positive Control DNA	IGH POS	IGH POS (+)		2	X	
Negative Control DNA	NGS NE	- 45µl	2	^		

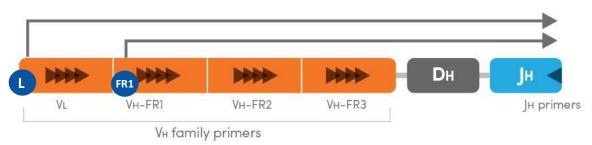


# LymphoTrack® SHM Assays



## Two primary options:

#### Schematic of the IGH Gene Locus:



## MiSeq® – 2 Options:

- IGHV Leader Somatic Hypermutation Assay
  - Primers target the IGHV Leader sequence
- IGH FR1 Assay
  - Primers target the IGH FR1

## <u>Ion Torrent</u>™ – 1 Option:

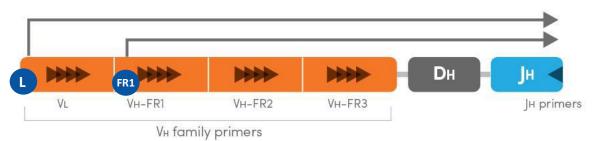
- IGH FR1 Assay
  - Primers target the IGH FR1



# LymphoTrack® Dx SHM Assays

## Two primary options:

#### Schematic of the IGH Gene Locus:



## MiSeq® – 2 Options:

- IGHV Leader Somatic Hypermutation Assay
  - Primers target the IGHV Leader sequence
- IGH FR1 Assay
  - > Primers target the IGH FR1

## **lon Torrent**™ – 1 Option:

- IGH FR1 Assay
  - Primers target the IGH FR1



# SHM LymphoTrack® Data



### LymphoTrack® IGH FR1 and IGHV Leader Somatic Hypermutation Assays

Sample Name

Total reads = 32,458

Easy identification of specific types of gene rearrangements such as IGHV3-21.

Rank	Sequence	Length	Merge count	V-gene	J-gene	% Total reads	Cumulative %	Mutation rate partial V-gene (%)	In-frame (Y/N)	No stop codon (Y/N)	V-coverage
1	TTCTCGTGGTG	455	29603	IGHV4-59_08	IGHJ4_02	9.93	9.93	11.26	Υ	Υ	98.63
2	CTCGCCCTCCT	463	205	IGHV5-51_01	IGHJ4_02	0.07	9.99	0.00	Υ	Υ	99.66
3	GGTTTTCCTTG	484	201	IGHV3-7_01	IGHJ4_02	0.07	10.06	7.77	Υ	Υ	100.00
4	CTCGCCCTCCT	463	185	IGHV5-51_01	IGHJ5_02	0.06	10.12	6.08	Υ	Υ	99.32
5	CTCGCCCTCCT	469	170	IGHV5-51_01	IGHJ4_02	0.06	10.18	0.00	Υ	Υ	99.32
6	CTCGCCCTCCT	466	160	IGHV5-51_01	IGHJ4_02	0.05	10.23	0.00	Υ	Υ	99.66
7	CTGCTGCTGAC	460	159	IGHV2-5_10	IGHJ5_02	0.05	10.29	8.08	Υ	Y	97.64
8	GGTTTTCCTTG	493	156	IGHV3-48_02	IGHJ6_02	0.05	10.34	3.72	Υ	Υ	98.99
9	CTCGCCCTCCT	334	153	IGHV5-51_02	IGHJ2_01	0.05	10.39	3.72	Υ	N	27.70
10	CTCGCCCTCCT	334	152	IGHV5-51_02	IGHJ2_01	0.05	10.44	3.38	Υ	N	26.01



# SHM LymphoTrack® Dx Data



### LymphoTrack® Dx IGH FR1 and IGHV Leader Somatic Hypermutation Assays

Sample Name

Total reads = 32,458

Easy identification of specific types of gene rearrangements such as IGHV3-21.

Rank	Sequence	Length	Merge count	V-gene	J-gene	% Total reads	Cumulative %	Mutation rate partial V-gene (%)	In-frame (Y/N)	No stop codon (Y/N)	V-coverage
1	TTCTCGTGGTG	455	29603	IGHV4-59_08	IGHJ4_02	9.93	9.93	11.26	Υ	Υ	98.63
2	CTCGCCCTCCT	463	205	IGHV5-51_01	IGHJ4_02	0.07	9.99	0.00	Υ	Υ	99.66
3	GGTTTTCCTTG	484	201	IGHV3-7_01	IGHJ4_02	0.07	10.06	7.77	Υ	Υ	100.00
4	CTCGCCCTCCT	463	185	IGHV5-51_01	IGHJ5_02	0.06	10.12	6.08	Υ	Υ	99.32
5	CTCGCCCTCCT	469	170	IGHV5-51_01	IGHJ4_02	0.06	10.18	0.00	Υ	Υ	99.32
6	CTCGCCCTCCT	466	160	IGHV5-51_01	IGHJ4_02	0.05	10.23	0.00	Υ	Υ	99.66
7	CTGCTGCTGAC	460	159	IGHV2-5_10	IGHJ5_02	0.05	10.29	8.08	Υ	Y	97.64
8	GGTTTTCCTTG	493	156	IGHV3-48_02	IGHJ6_02	0.05	10.34	3.72	Υ	Υ	98.99
9	CTCGCCCTCCT	334	153	IGHV5-51_02	IGHJ2_01	0.05	10.39	3.72	Υ	N	27.70
10	CTCGCCCTCCT	334	152	IGHV5-51_02	IGHJ2_01	0.05	10.44	3.38	Υ	N	26.01



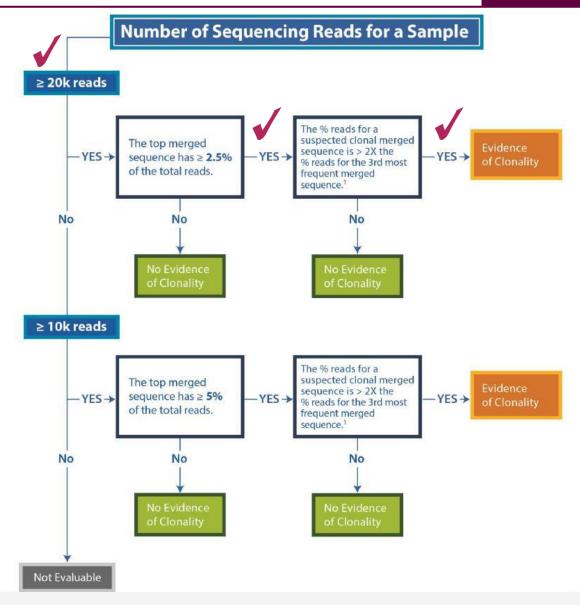
# Interpreting SHM – Step 1



## 1/2 - Determine Clonality

Total Read Count 474947

Rank	Sequence	Length	Merge count	V-gene	J-gene	% total reads	Cumulativ e %
1	ттстсстветвес	455	50248	IGHV4- 59_08	IGHJ4_02	10.58	10.58
2	CTGCTACTGACTG	319	192	IGHV2- 70_10	IGHJ4_02	0.04	10.62
3	CTGCTGCTGACCA	466	175	IGHV2- 5_01	IGHJ5_01	0.04	10.66
4	CTGCTGCTGACCA	457	162	IGHV2- 5_05	IGHJ6_02	0.03	10.69
5	CTGCTGCTGACCA	474	154	IGHV2- 5_05	IGHJ4_02	0.03	10.72
6	CTGCTGCTGACCA	454	150	IGHV2- 5_10	IGHJ5_02	0.03	10.76





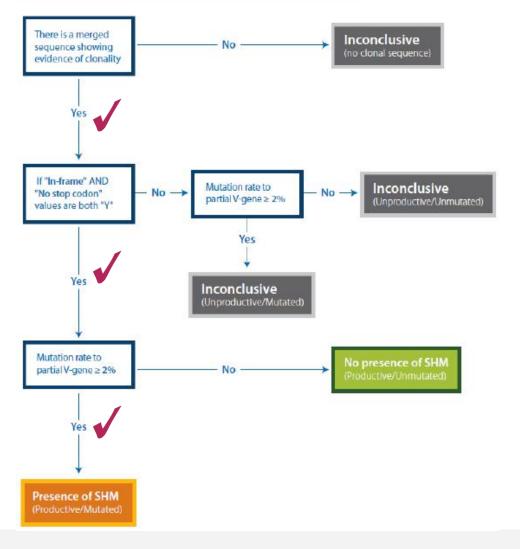
# Interpreting SHM – Step 2



## 2/2 - Determine SHM Status

Rank	Sequence	Length	Merge count	V-gene	J-gene	% total reads	Cumulativ e %	Mutation rate to partial V- gene (%)	In-frame (Y/N)	No Stop codon (Y/N)	V- coverage	CDR3 Seq
1	TTCTCGTGGTGGC	455	50248	IGHV4- 59_08	IGHJ4_02	10.58	10.58	11.26	Υ	Υ	98.63	GCGAGACGGAGC
2	CTGCTACTGACTG	319	192	IGHV2- 70_10	IGHJ4_02	0.04	10.62	4.32	n/a	N	35.55	not found
3	CTGCTGCTGACCA	466	175	IGHV2- 5_01	IGHJ5_01	0.04	10.66	6.62	Y	Y	100.00	GCACACAGACCG(
4	CTGCTGCTGACCA	457	162	IGHV2- 5_05	IGHJ6_02	0.03	10.69	2.99	Y	Y	99.67	GCACACAGATACT
5	CTGCTGCTGACCA	474	154	IGHV2- 5_05	IGHJ4_02	0.03	10.72	3.99	Y	Y	99.67	GCACACAGATACT
6	CTGCTGCTGACCA	454	150	IGHV2- 5_10	IGHJ5_02	0.03	10.76	11.78	Y	Y	98.99	GCATATGGTGTAA

#### **Suggested SHM Interpretation Criteria**





# Data Interpretation – IGHV and FR1



#### 1/2 - Determine Clonality

LymphoTrack Report for assay LEADER

Sample name: Leader\_positive\_S23\_L001\_001\_combined

**Ensure Total Read Count** is >20000 (or 10000)

Total Read Count: 474947

IndexQ30: 87 88

Caution: Do not edit fields and save.

2. Check Q30 Score

3. Top ranked sequence should account for at least 2.5% (or 5%) of total reads

4. Top ranked sequence should be >2x the 3rd ranked sequence

Top 10 Merged Read Summary

	Rank	Sequence	Length	Merge count	V-gene	J-gene	% total reads	Cumulativ e %	Mutation rate to partial V- gene (%)	In-frame (Y/N)	No Stop codon (Y/N)	V- coverage	CDR3 Seq
_	1	TTCTCGTGGTGGC	455	50248	IGHV4- 59_08	IGHJ4_02	10.58	10.58	11.26	Y	Y	98.63	GCGAGACGGAGC
	2	CTGCTACTGACTG	319	192	IGHV2- 70_10	IGHJ4_02	0.04	10.62	4.32	n/a	Ν	35.55	not found
1	3	CTGCTGCTGACCA	466	175	IGHV2- 5_01	IGHJ5_01	0.04	10.66	6.62	Y	Y	100.00	GCACACAGACCG
	4	CTGCTGCTGACCA	457	162	IGHV2- 5_05	IGHJ6_02	0.03	10.69	2.99	Y	Y	99.67	GCACACAGATACT
	5	CTGCTGCTGACCA	474	154	IGHV2- 5_05	IGHJ4_02	0.03	10.72	3.99	Y	Y	99.67	GCACACAGATACT
	6	CTGCTGCTGACCA	454	150	IGHV2- 5_10	IGHJ5_02	0.03	10.76	11.78	Y	Y	98.99	GCATATGGTGTAA
	7	CTGCTGCTGACCA	469	139	IGHV2- 5_01	IGHJ4_02	0.03	10.78	1.32	Y	Y	97.68	GCACTCGCGACAC
	8	стедесетесте	466	139	IGHV5- 51_01	IGHJ4_02	0.03	10.81	7.09	Y	Y	99.32	GCGAGATACTAT
	9	CTGCTACTGACTG	490	137	IGHV2- 70_10	IGHJ3_02	0.03	10.84	0.66	Y	Y		GCACGGATTCCTC
	10	CTGCTGCTGACCA	478	135	IGHV2- 5_10	IGHJ6_02	0.03	10.87	3.70	Y	Y		GCATACACTTGTT



# Data Interpretation – IGHV SHM

#### LymphoTrack Report for assay LEADER

Sample name: Leader\_positive\_S23\_L001\_001\_combined

Total Read Count: 474947

IndexQ30: 87.88

Caution: Do not edit fields and save.

Top 10 Merged Read Summary

Rank	Sequence	Length	Merge count	V-gene	J-gene	% total reads	Cumulativ e %	Mutation rate to partial V- gene (%)	In-frame (Y/N)	No Stop codon (Y/N)	V- coverage	CDR3 Seq
1	TTCTCGTGGTGGC	455	50248	IGHV4- 59_08	IGHJ4_02	10.58	10.58	11.26	Y	Y	98.63	GCGAGACGGAGC
2	CTGCTACTGACTG	319	192	IGHV2- 70_10	IGHJ4_02	0.04	10.62	4.32	n/a	1	35.55	not found
3	CTGCTGCTGACCA	466	175	IGHV2- 5_01	IGHJ5_01	0.04	10.66	6.62	Y	Y	100.00	GCACACAGACCG
4	CTGCTGCTGACCA	457	162	IGHV2- 5_05	IGHJ6_02	0.03	10.69	2.99	Y	Y	99.67	GCACACAGATACT
5	CTGCTGCTGACCA	474	154	IGHV2- 5_05	IGHJ4_02	0.03	10.72	3.99	Υ	Y	99.67	GCACACAGATACT
6	CTGCTGCTGACCA	454	150	IGHV2- 5_10	IGHJ5_02	0.03	10.76	11.78	Y	Y	98.99	GCATATGGTGTAA
7	CTGCTGCTGACCA	469	139	IGHV2- 5_01	IGHJ4_02	0.03	10.78	1.32	Y	Y	97.68	GCACTCGCGACAC
8	стедесетесте	466	139	IGHV5- 51_01	IGHJ4_02	0.03	10.81	7.09	Υ	Y	99.32	GCGAGATACTATT
	CTGCTACTGACTG		137	IGHV2- 70_10	IGHJ3_02	0.03	10.84	0.66	Y	Υ	99.34	GCACGGATTCCTG
10	CTGCTGCTGACCA	478	135	IGHV2- 5_10	IGHJ6_02	0.03	10.87	3.70	Y	Y	98.99	GCATACACTTGTT

#### 2/2 - Determine SHM Status

5. If sample is clonal, check whether clone is productive (Y+Y = Productive)

6. Identify whether mutation rate is >2%

Note: This analysis is also possible with *IGH FR1* 



# Case Study – Oxford Univ. Hospitals



Leukemia (2016), 1−9
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www.nature.com/leu

#### **ORIGINAL ARTICLE**

# Targeted deep sequencing reveals clinically relevant subclonal IgHV rearrangements in chronic lymphocytic leukemia

B Stamatopoulos<sup>1,2,3,7</sup>, A Timbs<sup>1,7</sup>, D Bruce<sup>1</sup>, T Smith<sup>4</sup>, R Clifford<sup>1,3</sup>, P Robbe<sup>1,3</sup>, A Burns<sup>1,3</sup>, DV Vavoulis<sup>3</sup>, L Lopez<sup>5</sup>, P Antoniou<sup>3</sup>, J Mason<sup>1</sup>, H Dreau<sup>1</sup> and A Schuh<sup>1,6</sup>

The immunoglobulin heavy-chain variable region gene (IgHV) mutational status is considered the gold standard of prognostication in chronic lymphocytic leukemia (CLL) and is currently determined by Sanger sequencing that allows the analysis of the major clone. Using next-generation sequencing (NGS), we sequenced the IgHV gene from two independent cohorts: (A) 270 consecutive patient samples obtained at diagnosis and (B) 227 patients from the UK ARCTIC-AdMIRe clinical trials. Using complementary DNA from purified CD19+CD5+ cells, we demonstrate the presence of multiple rearrangements in independent experiments and showed that 24.4% of CLL patients express multiple productive clonally unrelated IgHV rearrangements. On the basis of IgHV-NGS subclonal profiles, we defined five different categories: patients with (a) multiple hypermutated (M) clones, (b) 1 M clone, (c) a mix of M-unmutated (UM) clones, (d) 1 UM clone and (e) multiple UM clones. In population A, IgHV-NGS classification stratified patients into five different subgroups with median treatment-free survival (TFS) of > 280(a), 131(b), 94(c), 29(d), 15(e) months (P < 0.0001) and a median OS of > 397(a), 292(b), 196(c), 137(d) and 100(e) months (P < 0.0001). In population B, the poor prognosis of multiple UM patients was confirmed with a median TFS of 2 months (P = 0.0038). In conclusion, IgHV-NGS highlighted one quarter of CLL patients with multiple productive IgHV subclones and improves disease stratification and raises important questions concerning the pre-leukemic cellular origin of CLL.

Leukemia advance online publication, 9 December 2016; doi:10.1038/leu.2016.307

Stamatopoulos, B., Timbs, A., Bruce, D. et al. Targeted deep sequencing reveals clinically relevant subclonal IgHV rearrangements in chronic lymphocytic leukemia. *Leukemia* 31, 837–845 (2017). https://doi.org/10.1038/leu.2016.307



# Sample Types – 2 Independent Cohorts



## Cohort 1 (n=270)

- Consecutive subject samples obtained during baseline testing
- Peripheral blood, CD19+ cells were selected
- RNA extracted & cDNA were tested

## Cohort 2 (n=227)

- Subjects from the UK ARCTIC-AdMIRe clinical trials
- Peripheral blood, no selection
- gDNA tested



Stamatopoulos, B., Timbs, A., Bruce, D. et al. Targeted deep sequencing reveals clinically relevant subclonal IgHV rearrangements in chronic lymphocytic leukemia. *Leukemia* 31, 837–845 (2017). https://doi.org/10.1038/leu.2016.307





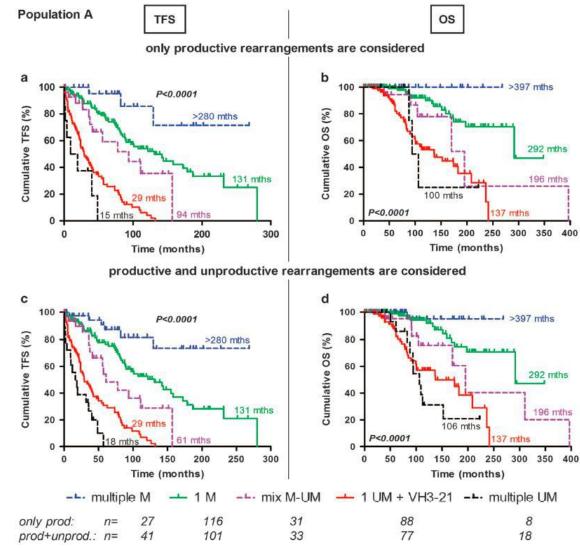
# In 24.4% of CLL samples tested multiple productive subclones are identified

2 studies for a total of 497 samples

## Five categories

- Multiple mutated clones
- 2. One mutated clone
- 3. Mix of mutated and unmutated clones
- 4. One unmutated clone, but the presence of a VH3-21 clone
- 5. Multiple unmutated clones

In this study the prognostic value of productive vs. unproductive rearrangements was the same



# NGS vs. Sanger Sequencing



## Sequence and mutation rate for most abundant clone compared

- IGHV mutation status concordance = 99.6% (235/236)
  - > Discordance in biclonal sample, small unmutated clone identified by Sanger
- Same sequence identified in 97.9% of samples (231/236)
  - > 5 cases the discordance was 1 bp and in 4 of these cases NGS sequence was closest to germline suggesting sequencing error with Sanger



## Conclusions

 IgHV mutational status by NGS refines IgHV Sanger Sequencing classification and can be used to define five different prognostic subgroups in an unselected population

 NGS-IgHV classification was able to precisely classify subjects with multiple IgHV rearrangements for which Sanger Sequencing was inconclusive and improved prognostication for 92 out of 270 cases

Stamatopoulos, B., Timbs, A., Bruce, D. et al. Targeted deep sequencing reveals clinically relevant subclonal IgHV rearrangements in chronic lymphocytic leukemia. *Leukemia* 31, 837–845 (2017). https://doi.org/10.1038/leu.2016.307



# Promotional Tools - Flyers



## LymphoTrack® Flyers (RUO) – MiSeq® and Ion S5/PGM<sup>TM</sup>



#### Portfolio for the MiSeq®

#### LymphoTrack Assays

LymphoTrack Assays represent a significant improvement over existing clonality assays as they efficiently detect the majority of B- and T-cell gene rearrangements and at the same time, identify the specific DNA sequence for each clonal gene

Therefore, these products have two important and complementary uses: aiding in the detection of initial clonal populations, and identifying sequence information required to track those clones in subsequent samples.

In addition, the LymphoTrack IGH and IGHV Leader Somatic Hypermutation Assays define the extent of somatic hypermutation present in the IGHV gene of analyzed samples

Our LymphoTrack multiplex master mixes are designed with Illumina® adapters and up to 48 indices. This allows for a one-step PCR and pooling of amplicons from several different samples and targets onto a single Illumina® MiSeq® flow cell.

Our LymphoTrack MRD Software allows for easy identification of residual clonotype sequences in subsequent samples.

- Identify, track, and assess mutation status of B- and T-cell gene rearrangements

Ordering	information
CATALOG #	PRODUCTS
7-121-0129	LymphoTrack* IGH FR1/2/3 Assay Kit A - MISeq*
7-121-0139	LymphoTrack* IGH FR1/2/3 Assay Panel - MiSeq*
7-121-0009	LymphoTrack* IGH FR1 Assay Kit A - MiSeq*
7-121-0039	LymphoTrack* IGH FR1 Assay Panel - MiSeq*
7-121-0149	LymphoTrack® IGH FR1 Assay Panel B - MiSeq®
7-121-0089	LymphoTrack* IGH FR2 Assay Kit A - MiSeq*
7-121-0099	LymphoTrack® IGH FR2 Assay Panel - MISeq®
7-121-0109	LymphoTrack® IGH FR3 Assay Kit A - MiSeq®
7-121-0119	LymphoTrack* IGH FR3 Assay Panel - MiSeq*
7-121-0059	LymphoTrack* IGHV Leader Somatic Hypermutation Assay Kit A - M
7-121-0069	LymphoTrack* IGHV Leader Somatic Hypermutation Assay Panel - I
7-122-0009	LymphoTrack® IGK Assay Kit A - MiSeq®
7-122-0019	LymphoTrack* IGK Assay Panel - MiSea*

LymphoTrack® TRB Assay Kit A - MiSea® LymphoTrack® TRB Assay Panel - MiSeq®

7-225-0019 LymphoTrack\* TRG Assay Kit A - MiSeq\* LymphoTrack\* TRG Assay Panel - MISeq\* 7-500-0009 LymphoTrack® Software - MiSea® LymphoTrack® MRD Software

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Indices 1-8 (5 sequencing reactions each) Indices 1-24 (5 sequencing reactions each) Indices 1-8 (5 sequencing reactions each) Indices 1-24 (5 sequencing reactions each) Indices 25-48 (5 sequencing reaction each) Indices 1-8 (5 sequencing reactions each) Indices 1-24 (5 sequencing reactions each)

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Indices 1-8 (5 sequencing reactions each)

indices 1-24 (5 sequencing reactions each)

Indices 1-8 (5 sequencing reactions each)

1 CD complimentary with purchase

1 CD complimentary with purchase

Indices 1-24 (5 sequencing reactions each)

#### LymphoTrack\*

lon S5™/ PGM™

#### Portfolio for the Ion S5<sup>™</sup>/PGM<sup>™</sup>

LymphoTrack Assays represent a significant improvement over existing clonality assays as they efficiently detect the majority of B- and T-cell gene rearrangements and at the same time, identify the specific DNA sequence for each clonal gene

Therefore, these products have two important and complementary uses: aiding in the detection of initial clonal populations, and identifying sequence information required to track those clones in subsequent samples.

In addition, the LymphoTrack IGH Assays defines the extent of somatic hypermutation present in the IGHV gene of analyzed samples

Our LymphoTrack multiplex master mixes are designed with Thermo Fisher® adapters and 12 indices. This allows for a one-step PCR and pooling of amplicons from several different samples and targets onto a single Ion S5/PGM™ sequencing chip,

Our LymphoTrack MRD Software allows for easy identification of residual clonotype sequences in subsequent samples.



#### **Key Benefits**

- One-step PCR for amplicon and library generation
- Identify, track, and assess mutation status of B- and T-cell gene rearrangements
- Sequence amplicons from any LymphoTrack kit together
- Included bioinformatics software for easy analysis and interpretation
- Same reagents for clonality, somatic hypermutation, and minimal residual disease (MRD) testing

#### Ordering Information

CATALOG #	PRODUCTS
7-121-0057	LymphoTrack® IGH FR1/2/3 Assay - S5/PGM™
7-121-0007	LymphoTrack* IGH FR1 Assay - \$5/PGM1M
7-121-0037	LymphoTrack* IGH FR2 Assay - S5/PGM™
7-121-0047	LymphoTrack® IGH FR3 Assay - S5/PGM™
7-122-0007	LymphoTrack® IGK Assay - S5/PGM™
7-227-0007	LymphoTrack® TRG Assay - S5/PGM™
7-500-0007	LymphoTrack* Software - S5/PGM <sup>TM</sup>
7-500-0008	LymphoTrack® MRD Software

12 Indices - 5 sequencing reactions each 12 Indices - 5 sequencing reactions each 12 Indices - 5 sequencing reactions each

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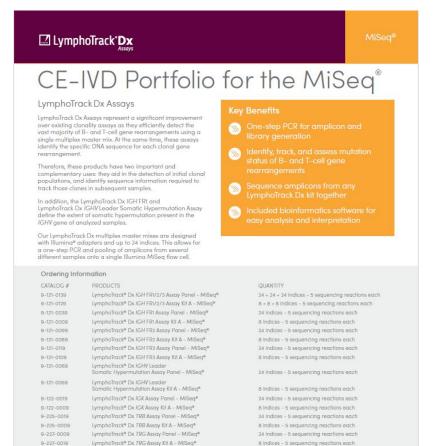
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# **Promotional Tools - Flyers**



## LymphoTrack® Flyers (CE-IVD) – MiSeq® and Ion S5/PGM<sup>TM</sup>



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LymphoTrack® Dx Software - MiSea®

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## **Promotional Tools - Postcards**



#### **Services Postcards**







# Take Home Message



- Invivoscribe offers solutions for SHM assessment for both Leader and FR1 assays
- The same NGS kits and software used for clonality can be used for SHM assessment
- NGS assays are easy, less time consuming and more sensitive than the traditional Sanger sequencing method





# Quiz

The IGH SHM Pos (+) Control DNA is included in all of the NGS kits that can be used to determine SHM status.

- True
- False

# What are the advantages of testing SHM on NGS platforms compared to testing with the gel method?

- Same kits than for clonality
- Easier, less time consuming, less labor intensive
- More sensitive
- All of the above







Somatic Hypermutation status needs to be assessed before determining Clonality.

- True
- False

The LymphoTrack<sup>®</sup> (Dx) software automatically calculates the SHM status by comparing the identified sequence with a germline sequence.

- True
- False







## Which assay(s) are available for the PGM and S5 platforms?

- LymphoTrack (Dx) IGH FR1 Assay
- LymphoTrack (Dx) IGHV Leader Assay
- All of the above

# Which products would you promote to a customer interested in assessing SHM status using FR1 primers on the MiSeq?

- LymphoTrack (Dx) IGH FR1 Assay MiSeq
- Lymphotrack (Dx) Software MiSeq
- IGH SHM Pos (+) Control DNA
- Specimen Control Size Ladder
- All of the above



